

Arising Model Organism Nervous System in *C. elegans* Biology: A Survey

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ABSTRACT

Although it's apparent simplicity, the nematode worm *C.elegans* has been a priceless tool for biological research, principally in the functional characterization of unique targets that have been identified the using genomics technologies. *C. elegans* and advanced organisms, along with the simplicity and cost-effectiveness of the agriculture, make for an effective in vivo model which is amenable to whole-organism high productivity compound screens and extensive target validation. This review shows how can be used *C. elegans* models to advance understanding of the molecular mechanisms of development and nervous system function.

Key words: *C. elegans*, life cycle, Genome, nervous system.

INTRODUCTION

Caenorhabditis elegans is a free-living (non- parasitic), transparent nematode (roundworm), unregimented, cylindrical body that tapers at both ends, 1 mm in length, that lives in temperate soil environments. The name is a mixture of Greek (caeno- - recent, rhabditis - rod-like) and Latin (elegans - elegant). First described by Emil Maupas in 1900¹. While *C. elegans* has been known and studied in laboratories nematologists for several years, it was not even Sydney Brenner in Cambridge, United Kingdom, chose this type of a new program in genetic research it has become a global phenomenon^{2,3}. Wanted species that was easy to keep it .That was tractable genetics (so you can isolate mutants and crosses made), and that it was easy to observe. Brenner has attracted a team of geneticists remarkable to join him, and beat *C. Elegans* researchers three Nobel Prizes for discoveries made through the new object model. *Caenorhabditis elegans* (*Caenorhabditis elegans*, hereinafter referred to as *C. elegans*) was first developed in 1963 by Sydney Brenner as a model organism for developmental biology and neurobiology research⁴.

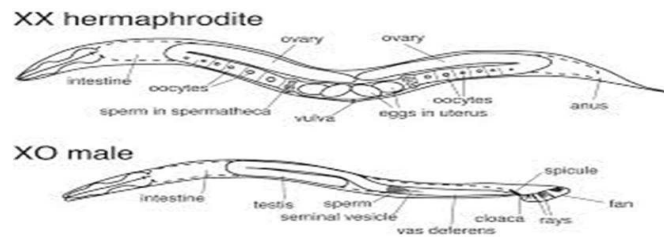
From the nematodes in the promotion of biotechnology in terms of: *C. elegans* cell lines identified widely used genetic techniques allow researchers to its produce great interest⁵. Researchers have used *C. elegans* as a research object in our knowledge of biological processes has made many important advances, such as development, apoptosis, and neurobiology gene regulation and aging and other fields⁶. But also as a very simple organisms, nematodes for the application of new technologies provides an ideal platform as the first to be sequenced⁷ multi-cellular organisms, the researchers first will be applied to chip technology^{8,9} and for the medical field of biomedical research provides broad prospects. For example, although the size of the genome of *C.elegans* genome of a human one-thirtieth, but it contains the majority of human gene homologues.

1-The reason for the study of nematodes *C. elegans*

From the structural characteristics of the nematode it is: Nematode sare small (length of about 1 mm), body transparent conducive experimental operation and observation, bacteria such as *E. coli* class for food, culture and inexpensive laboratory scale can be achieved culture. Nematodes have five pairs of autosomes and a **single pair** of sex chromosomes, two kinds of hermaphrodites and males (as shown in figure 1).

Nematode sand fruit fly to determine the gender of the same proportion of sex chromosomes and autosomes determine nematode sex. Hermaphrodite nematode can produce itself or with the male mating. In the laboratory, by the male or hermaphrodite self-hybridization with a specific manner to produce progeny phenotype, there by to facilitate genetic analysis. Further more, by selfing *C. elegans* hermaphrodite can produce about 300 to 350 progeny, mating with the male progeny will produce more of these characteristics is very conducive to the needs of a large number of genetic studies in phenotype and genotype. Nematode reproduction cycle of fast and short, generally only three days, the same as the time .

Fig.1: The two sexes of *C. elegans*



Wormbook.org

Required for yeast genetic hybridization; under suitable conditions for survival, nematode life cycle of about 2 to 3 weeks. Compared with other model organism *C. elegans*, the biggest advantage is its wild-type individual contains 959 cells, the location and number of cells is one to one. By the Waterston, *et al.*¹⁰ in the 1980s completed the full study investigated cell germ line genes influence cell fate mechanism provides a great convenience. We have nematodes by observing the *in vivo* cell division and cell migration in the process to determine the wild-type cell line¹¹. From the point of view of optical imaging, nematodes are very suitable for analysis by an optical microscope. Embryos (about 55 μm) or adult (1 mm) in size just within the range observed in the microscope, whereby we can observe the maximum numerical aperture of the embryo or larvae to adults and even a specific part of the body. Therefore, we can observe in the context of the whole animal study subcellular localization and other related information.

Nematodes study of the nervous system is concerned, each nematode cells (adult hermaphrodites were 959, adult male with 1031) has been drawn the developmental outcome of the pattern¹², these cell strains of the type of the individual no great difference between. Nematodes also has the most simple biological nervous system, in hermaphrodites, there are 302 neurons constitute the connection has been completely drawn into a clear pattern, by researchers called the image of "small-world" networks¹³. Researchers have found many such as chemo taxis, vigorous resistance, hydraulic conductivity, and male mating behavior of neural mechanisms such as nematodes.

2-Life cycles of *C. elegans*

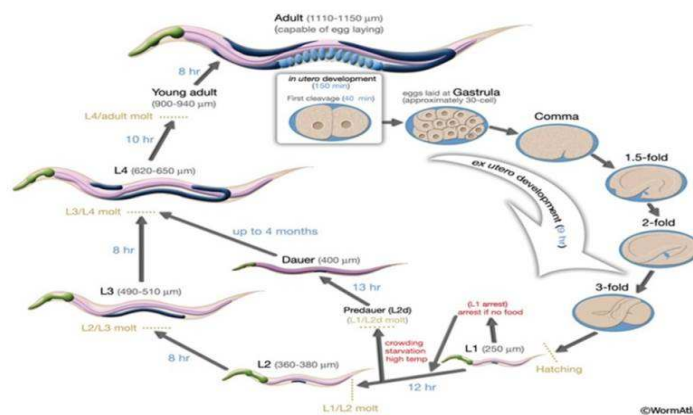
Have a short life cycle and production large numbers of progeny by sexual reproduction which occurs by self-fertilization in the hermaphrodite worms or mating with males? Self-fertilization allows worms homozygous for breeding and isolation of large real easy and maintenance of mutant strains. It is also a handy feature if paralyzed mutant animals because reproduction does not require movement in order to find and mate with males is necessary to move mutations between strains.

Hermaphrodite self-fertilized adult is capable of producing 300 progeny while male -fertilized hermaphrodites can produce more than 1,000 progeny .Each embryo to develop and hatch, and proceed through four larval stages (L1-L4) before becoming an adult¹⁴.

Under laboratory conditions the optimal average life span of *C. Elegans* 2-3 weeks. Rapid life cycle. At 25°C embryogenesis (from fertilization until hatching) occurs in 14 hours. Evolution happens postembryonic in four larval stages (L1-L4) which lasts a total of about 35 hours.

Under the deprivation of food, which is called alternative development path Dauer larva can be launched. Dauer larvae can survive for several months at this stage and be structural, metabolic and behavioral adaptations that increase the life span of up to 10 times and assist in dispersing animals to new habitats. When food becomes available, Dauer larvae feed and continue development to the adult stage. (Figure 2) shows from zero min are fertilization; the blue numbers along the arrows refer to the length of time the animal spends at a some stage. First cleavage Occurs at about 40 min. post-fertilization. Eggs are laid outside in about 150 min. post-fertilization and during gastrula stage. The length of the animal at both stage are marked next to the stage name in micrometers (μm).

Fig.2: Life cycle of *C.elegans*



www.wormatlas.org

4-Laboratory Culture of *C. elegans*

Culture of *C. elegans* in the laboratory is simple and easily and relatively inexpensive¹⁵. Because it is typically grown in Petri dishes on agar seeded with a lawn of *Escherichia coli* as a food source also it can be grown in mass quantities using culture strategies and fermenter-like these devices. Worm stocks are stored frozen in liquid nitrogen indefinitely with good viability. The ability to store *C. elegans* frozen dramatically simplifies culture strategies and reduces costs associated with handling and maintaining wild-type and mutant strains of the worm¹⁶.

5-The online information resources for *C. elegans*

Widely use internet and World Wide Web (WWW) in the scientific community the corresponding rapid development of hypertext browser programs have made a bewildering array of online resources available for biological researcher¹⁷.

In 1978 was created The Caenorhabditis Genetics Center (CGC) to support this rapidly evolving model system, and remains the only comprehensive resource that collects, maintains and distributes *C. elegans* breeds. Frozen stocks allow permanent storage of and consequent unlimited access to ancestral material, which makes *C. elegans* an attractive model for evolutionary biology. CGC works in closely collaboration with Worm Base, which imports all information on available strains and users can to locate these data through a specialized, and the search interface.

Other crucial task of CGC is the coordination of *C. elegans* labels. Historically, worm researchers have acceded to strict agreements that have ensured the uniformity of gene names, are derived from either the mutant phenotype or the molecular nature of the gene product.

This has virtually removed the confusions and clashes which are so common in other model organisms, and are proves that to be even more valuable today as automated phenotypic data extraction and literature analyses become the most common, the sites (as shown in table1 Box 1 and 2) listed constitute a basic set of the major electronic *C. elegans* resources on the Web .Each site, customized by its keepers, may contain links to many other more specific resources such as specialized programs written by *C.elegans* researcher, data search engines, electronic versions of standard print materials (e.g., the worm Breeders Gazette). Photographic images of worms, the information that is now verbatim at ones accessible requires only standard computer hardware, a graphical link to the internet, and a little of time¹⁸.

Table 1: the online information resources for *C. elegans* from Antoshechkin, & Sternberg, 2007¹⁸

Box 1 <i>Caenorhabditis elegans</i> online information resources	
<i>C. elegans</i> II	http://www.ncbi.nlm.nih.gov/books/bv.fcgi?call=bv.View..ShowSection&rid=ce2
WormBook	http://www.wormbook.org
WormClassroom	http://www.wormclassroom.org
WormAtlas	http://www.wormatlas.org
WormImage	http://www.wormimage.org
WormBase	http://www.wormbase.org
WormGenes	http://www.ncbi.nlm.nih.gov/IEB/Research/Acmby/index.html?worm
RNAiDB	http://www.rnai.org
PhenoBank	http://www.worm.mpi-cbg.de/phenobank2/cgi-bin/MenuPage.py
<i>C. elegans</i> RNAi Phenome Database	http://omicspace.riken.jp/Ce/rnai/jsp/index.jsp
InteractomeDB	http://vidal.dfci.harvard.edu/interactomedb
n-Browse	http://nematoda.bio.nyu.edu:8080/NBrowse/N-Browse.jsp
WormBase interaction browser	http://www.wormbase.org/db/seq/interaction_viewer
IntAct	http://www.ebi.ac.uk/intact/index.jsp
BioGrid	http://www.thebiogrid.org
EDGEdb	http://edgedb.umassmed.edu
NEXTDB	http://nematode.lab.nig.ac.jp/index.html
Hope Laboratory Expression Pattern Database	http://bgypc059.leeds.ac.uk/~web/databaseintro.htm
BC <i>C. elegans</i> Gene Expression Consortium	http://elegans.bcgsc.ca/perl/eprofile/index
Stanford Microarray Database	http://smd.stanford.edu/index.shtml
NCBI Gene Expression Omnibus	http://www.ncbi.nlm.nih.gov/projects/geo
EBI ArrayExpress	http://www.ebi.ac.uk/arrayexpress
<i>C. elegans</i> SAGE libraries	http://tock.bcgsc.bc.ca/cgi-bin/sage170
Structural Genomics of <i>C. elegans</i>	http://sgce.cbse.uab.edu/index.php
Protein Data Bank	http://www.pdb.org
NCBI PubMed	http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?DB=pubmed

Box 2 <i>Caenorhabditis elegans</i> online experimental resources	
<i>Caenorhabditis</i> Genetics Center (CGC)	http://biosci.umn.edu/CGC
CGC Strain Request	http://biosci.umn.edu/CGC/Strains/request.htm
CGC Nomenclature Guide	http://biosci.umn.edu/CGC/Nomenclature/nomenguid.htm
<i>C. elegans</i> Fosmids at Geneservice Ltd	http://www.geneservice.co.uk/products/clones/Celegans_Fos.jsp
BC GSC Fosmid Search	http://elegans.bcgsc.ca/perl/fosmid/CloneSearch
<i>C. elegans</i> Cosmids	http://www.its.caltech.edu/~wormbase/userguide/OtherResource/ObtainingReagents.html#elegans_genomic
<i>C. elegans</i> ESTs (NEXTDB)	http://nematode.lab.nig.ac.jp/index.html
<i>C. elegans</i> Gene Knockout Consortium	http://www.celeganskoconsortium.omrf.org
National BioResource Project of Japan	http://shigen.lab.nig.ac.jp/c.elegans/index.jsp
NemaGENETAG	http://elegans.imbb.forth.gr/nemagenetag
WorfDB	http://worfdb.dfci.harvard.edu
ORFeome at Geneservice	http://www.geneservice.co.uk/products/cdna/Celegans_ORF.jsp
ORFeome at OpenBiosystems	http://www.openbiosystems.com/GeneExpression/Non%2DMammalian/Worm/CelegansORFs/

6-Genome of *C. elegans*

C. elegans was the first multicellular organism to have a genome sequence perfectly. Sequence was published in 1998 although some small gaps existed, and was completed last gap by October 2002 .The *C. elegans* genome which is approximately 100 million base pairs long¹⁹. There are six chromosomes in *C. Elegans* five pairs of autosomes (chromosomes I, II, III, IV, and V) and sex chromosome X. Hermaphrodites have two chromosomes X (XX designated). Males have one X chromosome (designated XO); presence of only one chromosome instead of a pair called homozygous state. Can produce this case because of the loss of one X chromosome or by mating. Males cannot produce progeny of their own.

However, they can cross-fertilize hermaphrodites. It is commonly used in *C. Elegans* genes for making combinations and a mitochondrial genome. Gene density is about 1 gene / 5kb, (5 kilo-base pairs). Introns, or non-expressed sequences, are 26% of the genome. Intragenic, some large regions contain repeated DNA sequences. Are arranged many of the genes in the operons, which are polycistronic series that are transcribed together. *C. elegans* and other nematodes are among a small number of eukaryotes currently known to be operons, and these include trypanosomes, flatworms notably the trematode *Schistosoma mansoni*, and tunicate a primitive chordate *Oikopleura dioica*. It is believed that many of the organisms have these operons²⁰.

The genome contains about 20,470 protein-coding genes. Number of known RNA genes in the genome has increased significantly due to the 2006 discovery of a new class of 21U-RNA genes²¹ and is now believed that the genome contains more than 16,000 genes RNA, up from 1,300 a few, such as in 2005²². Scientific curators continue to evaluate a set of known genes: new gene predictions continue to be added, including incorrect modified or removed. In 2003, was determined the sequence of the genome of the nematode *C. Related briggsae* also, allowing researchers to study the comparative genomics of these organisms two²³. The sequence of the genome of the nematode more of the same sex, for example. *Remanei*, *C. Japonica* and *C. brenneri* are under study²⁴ across the whole genome shotgun technique, which is less complete and accurate than the "hierarchical" or clone-by-clone approach that was used in *C. elegans*. physical map of the *C. elegans* genome, which consisting of overlapping cosmid and YAC clones covering most of the six chromosomes, has been constructed in order to facilitate cloning of genes that have been placed on the genetic map. In addition, ongoing efforts to obtain expressed sequence tags (ESTs) and open reading frame sequence by (OSTs) provided for all *C. elegans* genes a wide range of cDNAs nematodes. All ~ 20,000 had been predicted open reading frames (ORFS) in expression profiling under many conditions using sophisticated technology²⁵.

In addition, the analysis of genetics, such as "mapping genes, epistasis, mutations and pleiotropy" and analysis of the genome, such as "chromosome organization, the repeated sequence and analysis of the promoter," intertwined.

In the genetic analysis, which is an attractive feature in the base of *C. Elegans* is the ease of generating mutations. Several chemical mutagens, particularly EMS (ethyl methane sulfonate) operate efficiently on a worm.

C. elegans is diploid, so very deleterious mutations can be induced and propagated without killing the animal. Because of the situation the main reproduction is hermaphrodite self-fertilization, so the effect of making any mutation homozygous can be examined automatically, as a result of Mendelian segregation.

Moreover, the availability of multiple mutations is important in providing not only a knockout mutations that completely remove the activity of the gene, but also partial or conditional mutations that allow for dissection of complex gene function. The vast majority of these mutations are recessive, associated with partial or complete loss of gene function, but also has been the establishment of many of the rare gain function of mutations, which often provided the basic tools for further investigation²⁶.

7-Nematodes nervous system

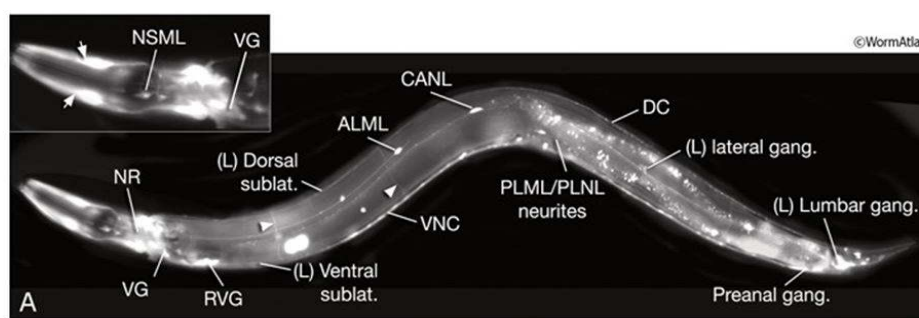
C. elegans nervous system is very simple, intersex adults' only 302 neurons, adult males because mating needs, more than the number of neurons in the adult sexes, reaching 381. Including a nerve ring, an abdominal nerve cord, and a back and a head nerve cord complex nervous system²⁷. Nerve ring nematode's brain can be considered, which is located in the head, including almost all of the neurons in the middle and most of the sensory neuron axons. Dorsal and ventral nerve cord from the nerve ring has been extended to the end, including a variety of motor neurons, abdominal nerve cord also contains information from many sensory neurons and intermediate neurons issued processes. In addition to the more important of these neurons, there are some small neurons located in other parts of the body. Males and females only 302 neurons, according to their morphological structure and characteristics are divided into 118 categories, these neurons through 5000 chemical synapses, 2000 neuromuscular junctions and 600 gap junctions interconnecting.

A typical neuron nematodes include a cell body, the cell body are called dendrites and two axons processes, branching dendrites generally do not responsible for smaller quantities from other neurons receiving synaptic input electron microscope postsynaptic structure is difficult to see, which increases the complexity of the synaptic connections. Axons are usually no bifurcation, responsible for synaptic output signals to a number of other neuronal and muscle. Typical of presynaptic structures can often be observed by an electron microscope, these features include the synaptic vesicle structure, dark matter in the presynaptic plasma membrane (darkening), and axon dark matter, and these materials must be provided for the storage vesicles space. Almost all of the synapses is in progress (en passant), and this indicates that the presynaptic axon becomes thicker and structures specialized storage spaces, and then narrowing distally extending^{28,29}. Similarly, dendritic synapses along its structure to accept the input signal is not terminated or the formation of specialized branches and other structures. Based on these structural and functional information, about half of the neurons neurons in the middle one-third of sensory neurons, one quarter of motor neurons (in addition there are some neurons may have more than one function).

Hermaphrodite nematodes has 302 neurons that belonging to two separate neurons of the nervous system: A large body of the nervous system (somatic nervous system), including the 282 neurons; are located in various ganglia in the head and tail and also along the ventral cord, the main longitudinal axon tract, 20 of these neurons are located inside the pharynx. Both nervous systems only through a pair of interconnected neurons in the middle RIP. Nematodes two nervous system topologies are quite different. Body nervous system neurons and their processes are generally located between the epidermis and body wall muscles through the basement membrane and muscle apart³⁰. Instead, throat neurons are located directly in the muscle of the pharynx; no base film of this structure exists. *C. elegans* nervous system classification is based on specific anatomical features, White *et al.*³¹. Based on electron microscopy observations of their topological structure and type of the synaptic connections of neurons into 118 categories, each category has a similar location, connection type axon synapses and dendrites³². Most of neuronal cell bodies concentrated in the head and tail ganglia³³.

C. elegans has 56, neuronal support cells (including the GLR cells, which supports only somatic nervous system connection. These neurons through about 6400 chemical synapses, 900 gap junctions (gap junction) and 1500 neuromuscular node (NMJs)^{34,35}. 75% of individual nematode chemical synapses can be regenerated. Name of each nematode neuron is composed of two or three capital letters, uppercase letters indicate which category belongs to neurons, and some names contain numbers, in order to distinguish the same categories of other neurons. If some neurons into radioactive symmetry (radially symmetrical), then these neurons will be named after the three capital letters plus L (on the left), R (on the right), D (back), V (ventral). Million to complete the list of nematode nerve, and their detailed description can be in the Individual Neuron Section Worm atlas^{31,36}. (As shown in figure 3)

Fig.3: The *C. elegans* nervous system



C. elegans nervous system is almost dispensable for its fertility, only CAN and M4 two very important neurons. So, for the multiple mutations in *C. elegans* nervous system can be separated out as fertile mutations. Since nematode body transparent, easy to operate observed under an optical microscope, and its cell structure is almost no change, any one or a group of neurons also are available through the laser burning³⁷ *C. elegans* neurons are divided into four types according to the functions: 1- Motor nerves (motor neurons), has a synapse with muscle cells connected .2- Sensory nerve (sensory neurons), with a clear perception of specialized functions.

3- Intermediate neurons (interneurons), receive incoming synaptic signals, the output signal reaches the synapse to spread to other neurons. 4- Multi-mode neurons (polymodal neurons), can perform the above functions. NSML / R are a multi-mode neuron is one pair of pharyngeal neurons, with improved secretion end, is classified as secretory nerve (also has motor function). In addition to the above categories, there are some neurons so far its function is still unknown. Some of which may guide the process and maintain a more important aspects of its role in the neural circuits rather than inside^{38,39}. for the nematode behavioral studies provide great convenience. In the larval stage of a few neurons can only lead to the loss of neurons in the nervous system function, without affect other functions of neurons can produce immeasurable damage, which allows us to examine a body or some neuron behavioral functions become possible. In addition, the application of technical records electrophysiological experiments nematode neurons and muscle activity of the technical problems have basically been solved, will greatly promote the *C. elegans* nervous system research fields and pace.

CONCLUSION

Small nematodes, *Caenorhabditis elegans*, and shares many of the structures and biological processes with more complex objects (such as humans). This, along with a short time to reproductive maturity (2-3 days), the period of life for two weeks, and detailed knowledge of our genetics have the function of each of the cells 959, Simple neuronal morphology makes it very easy to detect even minor developmental defects and make *C. elegans* organism's homepage scientists studying topics of the work of the nervous system. "This has helped" us to understand many aspects of human development, behavior, aging and diseases of the nervous system.

REFERENCES

1. Maupas, E. Modes et formes de reproduction des nematodes. Archives de Zoologie experimentale et generale, **8**: 463-624 (1901)
2. Brenner, S. The genetics of *Caenorhabditis elegans*. Genetics, **77(1)**: 71-94 (1974)
3. Brenner, S. Sequences and consequences. Philosophical Transactions of the Royal Society B: Biological Sciences, **365(1537)**: 207-212 (2010)
4. Hodgkin, J. Barnes, T.M. More is not better: brood size and population growth in a self-fertilizing nematode. Proceedings of the Royal Society of London Series B: Biological Sciences, **246(1315)**: 19-24 (1991)
5. Lai, H. J. Lo, S. J. Kage-Nakadai, E. Mitani, S. & Xue, D. The roles and acting mechanism of *Caenorhabditis elegans* DNase II genes in apoptotic DNA degradation and development. PloS one, **4(10)**: e7348 (2009)
6. Mitani, S. Neurogenesis in the nematode *C. elegans*. Tanpakushitsu Kakusan Koso, **38(15)**: 2534-2543 (1993)
7. Cutter, A. D. Dey, A. & Murray, R. L. Evolution of the *Caenorhabditis elegans* genome. Molecular biology and evolution, **26(6)**: 1199-1234 (2009)
8. Guha Thakurta, D. Palomar, L. Stormo, G.D. Tedesco, P. Johnson, T.E. Walker, D.W. & Link, C.D. Identification of a novel cis-regulatory element involved in the heat shock response in *Caenorhabditis elegans* using microarray gene expression and computational methods. Genome research, **12(5)**: 701-712 (2002)

9. Von Stetina, S.E. Watson, J.D. Fox, R.M. Olszewski, K.L. Spencer, W.C. Roy, P.J. & Miller, D.M. Cell-specific microarray profiling experiments reveal a comprehensive picture of gene expression in the *C. elegans* nervous system. *Genome biology*, **8(7)**: R135 (2007)
10. Waterston, R.H. Sulston, J.E. & Coulson, A.R. 2 the Genome. Cold Spring Harbor Monograph Archive, **33**: 23-45 (1997)
11. Hekimi, S. A neuron-specific antigen in *C. elegans* allows visualization of the entire nervous system. *Neuron*, **4(6)**: 855-865 (1990)
12. Greenwald, I. 19 Development of the Vulva. Cold Spring Harbor Monograph Archive, **33**: 519-541 (1997)
13. Siddiqui, S.S. & Culotti, J.G. Examination of neurons in wild type and mutants of *Caenorhabditis elegans* using antibodies to horseradish peroxidase. *Neurogenet*, **21(4)**: 271-289 (2007)
14. Tsang, W.Y. & Lemire, B.D. The role of mitochondria in the life of the nematode, *Caenorhabditis elegans*. *Biochimica ET Biophysica Acta (BBA)-Molecular Basis of Disease*, **1638(2)**: 91-105 (2003)
15. Lewis, J.A. & Fleming, J.T. Basic culture methods. In: *Caenorhabditis elegans: Modern Biological Analysis of an Organism*, (Epstein, H.F. and Shakes, D.C., eds.), Academic Press, New York, NY, 1995, pp. 4–29.
16. Strange, K. (Ed.). *C. elegans: methods and applications*, **351**: Springer (2006)
17. Riddle, D.L. Blumenthal, T. Meyer, J.R. *C.elegans II 1997* Cold Spring Harbor, NY Cold Spring Harbor Laboratory Press.
18. Antoshechkin, I. & Sternberg, P.W. The versatile worm: genetic and genomic resources for *Caenorhabditis elegans* research. *Nature Reviews Genetics*, July **8**:518-531 (2007)
19. Coghlan, A. Eichler, E.E. Oliver, S.G. Paterson, A.H. & Stein, L. Chromosome evolution in eukaryotes: a multi-kingdom perspective. *TRENDS in Genetics*, **21(12)**: 673-682 (2005)
20. Blumenthal, T. Operons in eukaryotes. *Briefings in functional genomics & proteomics*, **3(3)**: 199-211 (2004)
21. Ruby, J.G. Jan, C. Player, C. Axtell, M.J., Lee, W. Nusbaum, C. & Bartel, D.P. Large-Scale Sequencing Reveals 21U-RNAs and Additional MicroRNAs and Endogenous siRNAs in *C. elegans*. *Cell*, **127(6)**: 1193-1207 (2006)
22. Stricklin, S.L. Griffiths-Jones, S. & Eddy, S. R. (2005). *C. elegans* noncoding RNA genes.
23. Stein, L.D. Bao, Z. Blasiar, D. Blumenthal, T. Brent, M.R. Chen, N. & Waterston, R.H. The genome sequence of *Caenorhabditis briggsae*: a platform for comparative genomics. *PLoS biology*, **1(2)**: e45 (2003)
24. Genome Sequencing Center. "Caenorhabditis brenneri: Background". Washington University School of Medicine. Retrieved 2008-07-11.
25. Syntichaki, P. & N. Tavernarakis. Genetic Models of Mechanotransduction: The Nematode *Caenorhabditis elegans*. *Physiol Rev*, **84**: 1097–1153 (2004)
26. Hodgkin, J. (2005). Introduction to genetics and genomics.
27. Mitani, S. Neurogenesis in the nematode *C. elegans*. *Tanpakushitsu Kakusan Koso*, **38(15)**: 2534-2543 (1993)
28. Zetka, M.C. Kawasaki, I. Strome, S. & Müller, F. Synapsis and chiasma formation in *Caenorhabditis elegans* require HIM-3, a meiotic chromosome core component that functions in chromosome segregation. *Genes & development*, **13(17)**: 2258-2270 (1999)
29. MacQueen, A.J. Synapsis-dependent and -independent mechanisms stabilize homolog pairing during meiotic prophase in *C. elegans*. *Genes Dev*, **16(18)**: 2428-2442 (2002)
30. Karimi, M. Goldie, L.C. Ulgiati, D. & Abraham, L.J. Integration site-specific transcriptional reporter gene analysis using Flp recombinase targeted cell lines. *Bio Techniques*, **42(2)**: 217 (2007)
31. White, J.G. Southgate, E. Thomson, J.N. & Brenner, S. The structure of the nervous system of the nematode *Caenorhabditis elegans*. *Philosophical Transactions of the Royal Society of London. B, Biological Sciences*, **314(1165)** 1-340 (1986)

32. Bargmann, C.I. Neurobiology of the *Caenorhabditis elegans* genome. *Science*, **282(5396)**: 2028-2033 (1998)
33. Toth, M.L. Melentijevic, I. Shah, L. Bhatia, A. Lu, K. Talwar, A. & Driscoll, M. Neurite sprouting and synapse deterioration in the aging *Caenorhabditis elegans* nervous system. *The Journal of Neuroscience*, **32(26)**: 8778-8790 (2012)
34. Majewska, A. & Yuste, R. Topology of Gap Junction Networks in *C. Elegans*, *Journal of theoretical biology*, **212(2)**: 155-167 (2001)
35. Altun, Z.F. Chen, B. Wang, Z.W. & Hall, D.H. High resolution map of *Caenorhabditis elegans* gap junction proteins. *Developmental Dynamics*, **238(8)**: 1936-1950 (2009)
36. Hall, D.H. & Russell, R.L. The posterior nervous system of the nematode *Caenorhabditis elegans*: serial reconstruction of identified neurons and complete pattern of synaptic interactions. *The Journal of neuroscience*, **11(1)**: 1-22 (1991)
37. Fishman, T. & Hardy, A. Effect of spatial hole burning on injection-locked vertical-cavity surface-emitting laser arrays. *Applied optics*, **39(18)**: 3108-3114 (2000)
38. Chen, B.L. Hall D.H. Chklovskii, D.B. Wiring optimization can relate neuronal structure and function [J]. *Proc Natl Acad Sci* 103(12): 4723-4728 (2006)
39. Hall, D.H. Lints, R. Altun, Z. Nematode neurons: anatomy and anatomical methods in *Caenorhabditis elegans*[J]. *Int Rev Neurobiol*, **69**: 1-35 (2006)